

Effect of host plants on the functional response of the acarophagous ladybird beetle, *Stethorus gilvifrons* to *Tetranychus turkestan*

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Abstract

The strawberry spider mite, *Tetranychus turkestan* Ugarov and Nikolski, is an important pest of various field and glasshouse crops in Iran. This study aimed to determine the functional response of adult females of *Stethorus gilvifrons* Mulsant to different densities of *T. turkestan* on three fabaceous host plant species: cowpea (*Vigna unguiculata* L.), white bean (*Phaseolus lunatus* L.), and red bean (*Phaseolus calcaratus* Roxb.) at 30 ± 2 °C, 60 ± 10 % RH, and a photoperiod of 16L: 8D. Beetles were starved for 24 h before testing and then isolated singly for 24 h in 9-cm Petri dishes with 4, 8, 12, 24, 48, or 96 adult females of *T. turkestan*. Logistic regression analysis revealed a type III response on cowpeas. Constant (*b*) and handling time were 0.00718 and 0.3299 h, respectively. In contrast, a type II response was observed in white beans and red beans. The maximum attack (0.1219 h^{-1}) and the minimum handling time (0.2301 h) were recorded on white beans and red beans, respectively. The maximum attack rates (*T/Th*) were 72.74, 60.42, and 104.30 eggs/day on cowpeas, white beans, and red beans, respectively. This laboratory study suggests that the plant features affect the capability of *S. gilvifrons* to respond to changes in a high *T. turkestan* density.

Keywords: Attack rate, Handling time, Fabaceae, Strawberry spider mite

Introduction

The strawberry spider mite, *Tetranychus turkestanii* Ugarov and Nikolski (*Tetranychus atlanticus* McGregor), is among the most important tetranychid pests in the world (Jeppson et al., 1975; Bolland et al., 1998). It is a polyphagous species that feeds on 25 plant species belonging to more than 15 plant families (Jeppson et al., 1975; Kamali et al., 2001; Modarres Awal, 2012). This phytophagous mite is distributed throughout the United States, Europe, Russia, Japan, China, the Near East, and the Middle Eastern countries (Jeppson et al., 1975; Helle and Sabelis, 1985; Zhang, 2003; Hoy, 2011).

In Khuzestan province, southwest of Iran, *T. turkestanii* frequently attacks plant species belonging to the families, such as Solanaceae (Nemati et al., 2005), Fabaceae (Leguminosae) (Sohrabi and Shishehbor, 2008), and Cucurbitaceae (Karami Jamour et al., 2012). The feeding activity of *T. turkestanii* leads to the appearance of white chlorotic spots on the upper side of leaves with copious webbing underside the leaves. In serious infestations, leaves may drop, and flowering may be noticeably reduced. The short developmental period and high reproductive potential help the insect to have a fast-growing population, allowing the mite to incur economic losses in favorable conditions, resulting in a rapid decline of crop yields (Carey and Bradley, 1982; Tomczyk and Kropczynska, 1985; Sohrabi and Shishehbor, 2008; Karami Jamour et al., 2012).

Stethorus gilvifrons Mulsant (Coleoptera: Coccinellidae) is a coccinellid beetle native to Iran (Mossadegh and Kocheili, 1993) and several other countries in the east of Asia (McMurtry et al., 1970; Chazeau, 1985; Aksit et al., 2007). It is commonly found on field crops, such as sugarcane and castor bean (Afshari, 1999; Modarres Awal, 2012), and on fruit trees, such as apple, almond, cherry, pistachio, fig, apricot, lime trees, and date palms (Kohpayezadeh Isfahani and Mossadegh, 1993; Mossadegh and Kocheili, 1993; Mossadegh and Kocheili, 2003). *S. gilvifrons* has been validated as a good candidate for the biological control of numerous species of spider mites,

including *T. turkestanii* (Chazeau 1985; Biddinger et al., 2009; Hoy, 2011).

The performance of a predator depends on several characteristics, two of which are functional and numerical responses. The functional response describes the relationship between the number of preys attacked by an individual predator and prey density (Solomon, 1949). Several types of functional response have been described, including a linear increase (Type I), an increase decelerating to a plateau (type II), or a sigmoid increase (Type III) (Holling, 1959; Hassell et al., 1977). Arthropods feature type II and III responses, though type II is more frequent (Hassell, 1978).

The effect of host plant morphological characteristics on the predatory behavior of *Stethorus* species has been reported frequently (Putman, 1955; Riddick et al., 2011; Riddick and Wu, 2011). The information on the impact of Fabaceae host plants on the functional response of *S. gilvifrons* to *T. turkestanii* is limited. In fact, the only documented report was that of Karami Jamour and Shishehbor (2012), who reported that the functional response of *S. gilvifrons* on *T. turkestanii* varied among three Fabaceae plant species (cowpea, green gram, and pinto bean). To further quantify the influence of host plants on the predatory characteristics of *S. gilvifrons*, we determined the functional response of adult females to different densities of *T. turkestanii* on three Fabaceae host plant species.

Materials and methods

Plant materials

Three plant species of Fabaceae were used in this study: cowpea (*Vigna unguiculata* L.), white bean (*Phaseolus lunatus* L.), and red bean (*Phaseolus calcaratus* Roxb.). Seeds were purchased from a local grocery store and planted in plastic pots (20 cm diameter, 10 cm height) in a mixture of compost (Nutrifood Company, Iran) and sawdust (1:1 ratio) and kept in wooden cages (120 × 60 × 60 cm³) covered by a white nylon mesh (with apertures of 120 μm). The plants were maintained in the laboratory at 30±2 °C, 65± 5% RH, and 16:8 (L:D) with illumination (4000 lux)

provided by fluorescent lamps. The plants were watered every three days, and fertilizer (Hortigrow, Netherland) was added to the water. Studies were initiated when cowpeas, white beans, and red beans were three, four, and four weeks old, respectively. The heights of the above plants at these ages were ≈ 30 , 25, and 25 cm for cowpeas, white beans, and red beans, respectively. Only the second to fourth fully expanded leaves (from the growing terminal) were selected.

Spider mite colony

Tetranychus turkestanii was collected from morning glory (*Convolvulus arvensis* L.) leaves grown in the vicinity of the Shahid Chamran University of Ahvaz, Ahvaz, Iran (31° 20' N, 48° 40' E) in November 2018 and used to start rearing the colony. This stock colony was separately maintained on cowpea, white bean, and red bean plants. The infested plants were kept in wooden framed cages under the laboratory conditions described above. The plants were maintained until they were severely damaged by the spider mites. Also, new plants were added when necessary.

S. gilvifrons colony

Stethorus gilvifrons were collected from hollyhock (*Altha officinalis* L.) leaves grown in the vicinity of Shahid Chamran University of Ahvaz, Ahvaz, Iran, in April 2018 and were used to establish a stock colony. The colony was maintained in the above-mentioned wooden rearing cages using all stages of *T. turkestanii* as a food source on cowpea plants in the laboratory. New *T. turkestanii*-infested cowpea plants were added to the cages every week. The laboratory conditions were similar to those described above. The experiments were carried out at the Department of Plant Protection, Shahid Chamran University of Ahvaz, Ahvaz, Iran.

The leaves of cowpea, white bean, and red bean were used as substrates. The experimental arena consisted of a plastic petri dish (9 cm in diameter), the lid of which was covered with a fine mesh screen. Each petri dish was prepared by placing a leaf (upside down) of the respected plant on a wet cotton pad of the same size at the bottom of the petri dish. Adult female strawberry

spider mites (n=4, 8, 12, 24, 48, or 96) were placed on the leaves using a fine paintbrush. These densities were selected based on the preliminary tests on the consumption capacity of different stages of *S. gilvifrons*. A single female *S. gilvifrons* (approximately two weeks old), starved for 24 h before testing, was introduced into each petri dish. Ten replications of each density were set up simultaneously. The experiments lasted for 24 h in a growth chamber at 30±2 °C, 65±5 % RH, and 16: 8 (L:D), after which the beetles were removed, and the total number of consumed prey was counted. If the mite bodies were shrunk, discolored, or immobile, they would be registered as a consumed prey. Observations were made with a stereomicroscope.

The trichome numbers in the lower surface of plant species were counted using a dissecting microscope (Olympus Co, Japan) at 40X magnification. Trichomes were counted in an area of 1.0 mm² from different parts of each leaf.

Data analysis

A two-way factorial analysis of variance (ANOVA) and Tukey's honestly significant difference (HSD) tests ($P < 0.05$) were used to analyze the data for significant differences in the number of consumed prey and number of trichomes among the tested plant species (SAS Institute 2012).

The logistic regression of the proportion of the prey consumed as a function of initial prey density was used to determine the type of the functional response curve of *S. gilvifrons* for different densities of *T. turkestanii* on different host plant species:

$$\frac{N_a}{N_0} = \frac{\exp(P_0 + P_1 N_0 + P_2 N_0^2 + P_3 N_0^3)}{1 + \exp(P_0 + P_1 N_0 + P_2 N_0^2 + P_3 N_0^3)}$$

where N_a is the number of prey consumed, N_0 is the initial prey density, (N_a/N_0) is the probability of prey consumption, and P_0 , P_1 , P_2 , and P_3 are the estimated maximum likelihood of the constant, linear, quadratic, and cubic coefficients, respectively, related to the curve slope (Juliano, 2001). If the linear parameter P_1 is negative, a type II functional response will be evident, whereas P_1 is positive and P_2 is negative, and a

type III response is evident (Juliano, 2001). After determining the type of functional response, the attack rate (a) and handling time (T_h) coefficients were estimated (a nonlinear least-squares regression (PROC NLIN)). In the current study, we used an explicit deterministic model for type II functional response (Rogers, 1972):

$$N_a = N_0 [1 - \exp(-a(T_h N_a - T))] \quad (2)$$

where N_a is the number of prey killed, N_0 is the initial prey density, T is the total time that predator and prey are exposed to each other (24 h), a is the attack rate, and T_h is the handling time. For the type III functional response, in the simplest generalized form, the attack rate (equation 3) is a function of the initial quantity of prey:

$$a = (d + bN_0)/(1 + cN_0) \quad (3)$$

where b , c , and d are constants that must be estimated. The simplest form arises when a is a function of the initial density as in equation 4:

$$N_a = N_0 \{1 - \exp[(d + bN_0)(T_h N_a - T)(1 - cN_0)]\} \quad (4)$$

Then, for each host density, the attack rate coefficient (a) can be determined as $a = bN_0$. An iterative nonlinear least square regression (NLIN procedure, SAS Institute, 2012) was used to fit the random predator equation to the data to estimate the parameters describing the type II functional response. Likewise, a nonlinear least square regression procedure was used to fit the related equation to the data generating a type III functional response.

The parameters D_{Th} and D_a show differences between the two groups with the same functional response type in handling times and attack rates, respectively. After analysis using an indicator variable method as follows (Juliano, 2001; Farhadi et al., 2010; Zarghami et al., 2016):

$$N_a = \{1 - \exp[-(a + D_a(j))(T - (T_h + D_{Th}(j))N_a)]\}$$

Where j is an indicator variable that takes on a value of 0 for the first data set and 1 for the second data set. Testing the null hypothesis in that $D_a = 0$ reveals the significance of differences in searching efficiencies between the two stages (Juliano, 2001, Allahyari et al., 2004;

Farhadi et al., 2010; Zarghami et al., 2016). The coefficient of determination was calculated according to the following formula (Allahyari et al., 2004):

$$r^2 = 1 - \frac{\text{residual sum of squares/corrected total sum of squares}}$$

Results

The prey density and host plant had a significant effect on the mean number of the prey consumed (prey density: $F = 223.62$; $df = 5, 153$; $P < 0.0001$; host plant: $F = 10.57$; $df = 2, 153$; $P < 0.0001$), as did the prey density \times host plant interaction ($F = 3.54$; $df = 10, 153$; $P = 0.0003$) indicated that the effect of prey density treatment on the mean number of the prey consumed was affected by the host plant (Table 1).

The plant species evaluated in this work significantly varied in the density of trichomes ($F = 17.92$; $df = 2, 44$; $P < 0.0001$). If arranged in descending order, the plants with higher trichome density were white beans (42.93 ± 7.45), red beans (29.86 ± 4.76), and cowpeas (0.66 ± 0.23). The results indicated that *S. gilvifrons* could consume more adult female *T. turkestanii* on cowpeas than on white beans and red beans. In prey densities below 24, there was a small difference in the mean number of the prey consumed on the three host plants; however, the differences in the mean number of the prey consumed only became apparent at higher prey densities (Table 1). At the highest density of 128 adult females, the consumption rate was approximately 20 and 30 % higher on cowpeas than on red beans and white beans, respectively.

Functional response curves of *S. gilvifrons* on different host plant species are shown in Figures 1 & 2. Logistic regression analysis (Table 2) indicated a type II functional response on red beans and white beans but a type III response on cowpeas.

The results indicated that despite similar functional responses of *S. gilvifrons* feeding on different densities of adult female *T. turkestanii* on white beans and red beans (Type II functional response), the attack rate and handling

Table 1. Mean (\pm SE) (Range) of *Tetranychus turkestanii* adults attacked by *Stethorus gilvifrons* adult females on leaf with different prey density over a 24-h period at $30\pm^{\circ}$ C.

Prey density	Cowpea	White bean	Red bean	F	df	P
4	3.91 \pm 0.1 ^{Da}	3.8 \pm 0.13 ^{Da}	3.7 \pm 0.15 ^{Da}	0.58	2	0.56
8	8.00 \pm 0.1 ^{Da}	7.5 \pm 0.26 ^{Dab}	7.2 \pm 0.24 ^{CDb}	3.64	2	0.04
12	11.5 \pm 0.3 ^{Da}	11.7 \pm 0.15 ^{Da}	11.4 \pm 0.22 ^{CDa}	0.42	2	0.66
24	23.7 \pm 0.21 ^{Ca}	21.0 \pm 0.85 ^{Ca}	17.4 \pm 1.28 ^{Cb}	11.79	2	<0.0001
48	42.0 \pm 1.46 ^{Ba}	33.7 \pm 1.23 ^{Bb}	32.1 \pm 1.26 ^{Bb}	16.05	2	<0.0001
96	67.6 \pm 3.03 ^{Aa}	46.9 \pm 4.73 ^{Ab}	53.5 \pm 6.48 ^{ABb}	3.52	2	<0.0001
F	126.97	67.59	46.41			
df	5	5	5			
P	<0.0001	<0.0001	<0.0001			

Means within columns and rows followed by the same letters are not significantly different ($P < 0.05$). Upper case letters are used to compare the effects of different densities on the same host plant; lowercase letters are used to compare effects of different host plants on the same prey density.

time parameters were significantly different on the two host plants (Tables 3 and 4).

For cowpeas, the relationships between the attack rate and the initial number of preys were linear ($a=bN_0$), and the rate of successful attack (a) for densities of 4, 8, 12, 24, 48, and 96 were 0.0287, 0.0574, 0.0861, 0.1723, 0.3446, and 0.6893, respectively. According to the handling time estimation, the maximum number of *T. turkestanii* adult females that a single female of *S. gilvifrons* can predate is 60.42, 72.74, and 104.30 on the white bean, cowpea, and red bean as a host plant, respectively (Table 3).

Discussion

Variable results have been reported regarding the type of functional response of *S. gilvifrons* feeding on different tetranychid mites. Type III functional response of *S. gilvifrons* was reported by Hajizadeh et al. (1993) on *Panonychus ulmi* (Koch) and by Sohrabi and Shishehbor (2007) on *T. turkestanii*, which are in agreement with our findings on cowpea. Furthermore, other studies have also shown a type II functional response of *S. gilvifrons* to variations in densities of its prey, including *Oligonychus sacchari* McGregor (Afshari, 1999), *Tetranychus urticae* Koch (Mehr-Khou, 2006), *Oligonychus afraziaticus* McGregor (Matin,

2007), *Eutetranychus orientalis* Klein (Imani et al., 2010) and *T. turkestanii* (Karami Jamour and Shishehbor, 2012) which are similar to our results on white bean and red bean.

Holling (1965) reports that only predators exhibiting a type III functional response can regulate their prey populations because only this type of response will pave the way for long-term persistence. However, it should be noted that although the functional response is a significant aspect of natural enemy behavior, it is not the sole factor determining the effectiveness of a biological control agent. Other aspects, including numerical response, high intrinsic rate of population increase, high searching efficiency (Hassell, 1978), prey stage (Haque et al., 2010), age of natural enemy (Li et al., 2007), generation of a natural enemy (Khanamani et al., 2017), host plant characteristics (Cedola et al., 2001; Skirvin and Fenlon, 2001; Prevez and Omarkar, 2005; Ahn et al., 2010; Gontijo et al., 2012; Karami Jamour and Shishehbor, 2012), and abiotic environmental factors, such as temperature (Skirvin and Fenlon, 2003) can also affect the efficiency of predators. Although the only factor of the host plant species evaluated in the current study was trichome density, it is well-known that predatory effectiveness is affected by other plant traits. For instance, the

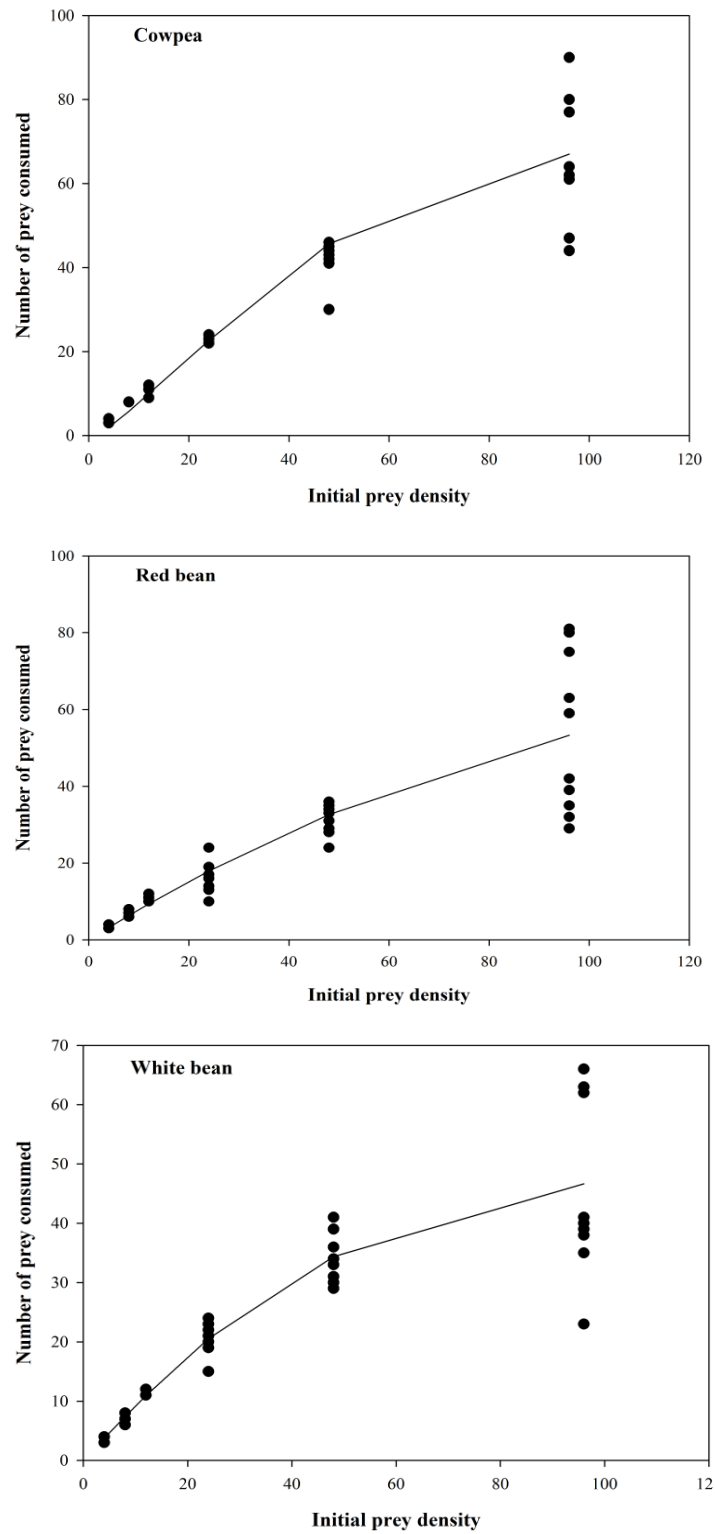


Figure 1. Functional response of *Stethorus gilvifrons* to different densities of *Tetranychus turkestani* on cowpea, white bean and red bean. Symbols are observed data and lines were predicted by model.

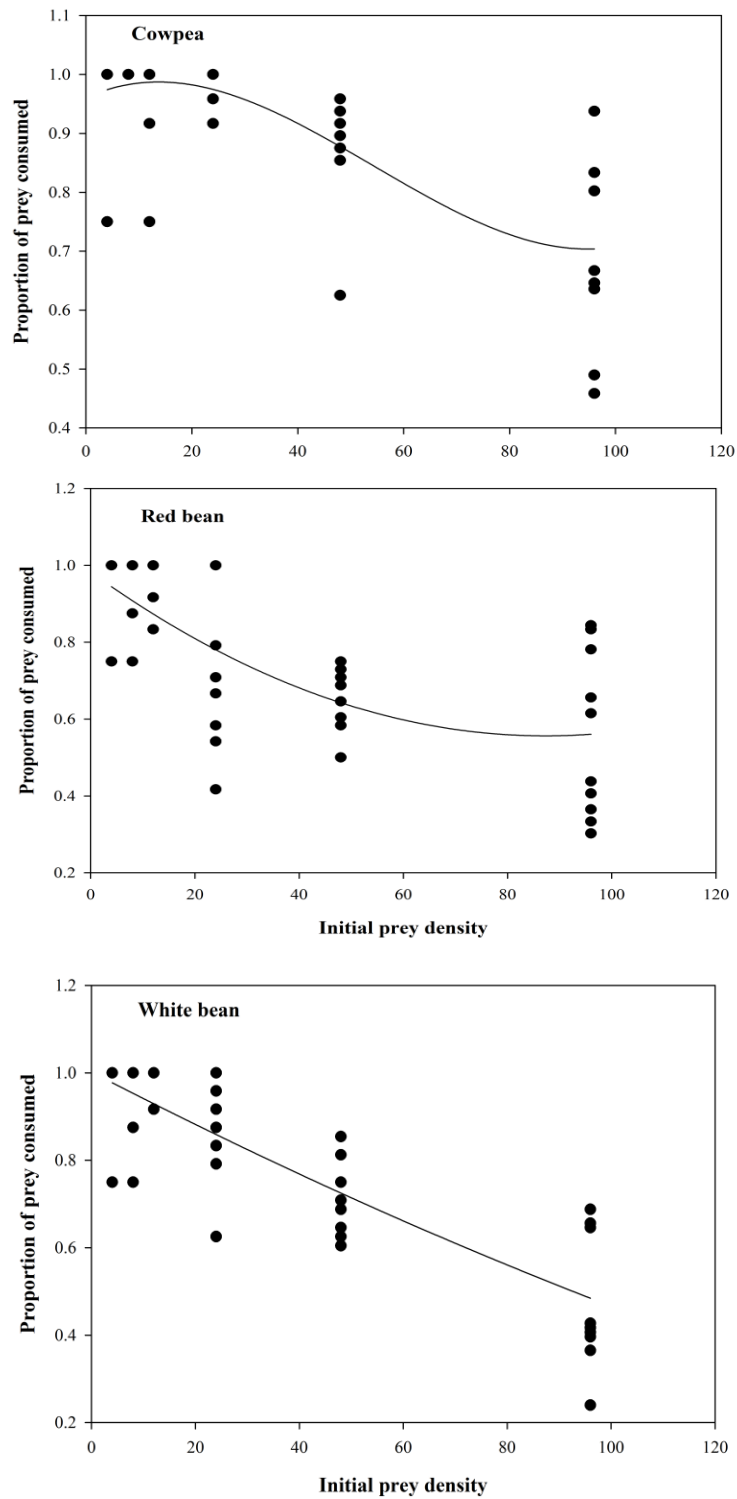


Figure 2: Proportion of *Tetranychus turkestani* consumed by *Stethorus gilvifrons* in relation to prey density on cowpea, white bean and red bean. Symbols are observed data and lines were predicted by model.

Table 2. Parameters (Mean±SE) for linear coefficient of the logistic model fit to proportion of prey density by adult female of *Stethorus gilvifrons* on different plants.

Host plant	Coefficient	Estimate	SE	χ^2	P
Cowpea	Intercept	3.0055	1.0378	8.39	0.0038
	Linear	0.1450	0.1095	1.75	0.1854
	Quadratic	-0.00521	0.0027	3.66	0.0559
	Cubic	0.000036	0.00002	4.19	0.0406
White bean	Intercept	3.8196	0.7536	25.69	<0.0001
	Linear	-0.0873	0.0670	1.70	0.1925
	Quadratic	0.00057	0.0016	0.13	0.7161
	Cubic	-8.45E-7	9.943E-6	0.01	0.9323
Red bean	Intercept	4.1260	0.5984	47.54	<0.0001
	Linear	-0.2207	0.0523	17.82	<0.0001
	Quadratic	0.00434	0.00122	12.64	0.0004
	Cubic	-0.00003	7.735E-6	11.01	0.0009

Table 3. Parameters (Mean±SE) estimated by Rogers model, evaluating functional response of *Stethorus gilvifrons* adult females to densities of *Tetranychus turkestanii* adults on different plants.

Plant	Type	a (Type II) or b (Type III) (h^{-1})	$T_h(h)$	T/ T_h	R ²
Cowpea	III	0.00718±0.00214	0.3299±0.0158	72.74	0.9601
White bean	II	0.1219±0.0301	0.3972±0.0428	60.42	0.944
Red bean	II	0.0691±0.0176	0.2301±0.0688	104.30	0.904

Table 4. Significant differences between functional response of *Stethorus gilvifrons* adult female fed on adult female *Tetranychus turkestanii* on white bean and red bean.

Parameter	Estimation	Asymptotic SE	Asymptotic 95% CI	
			Upper	Lower
D_a^*	0.0528	0.0392	0.1305	-0.0248
D_{Th}^{**}	0.1671	0.0791	0.3238	0.0105

* Indicating differences between treatments for a .

** Indicating differences between treatments for T_h .

The species of the host plant altered the functional response of *S. gilvifrons* with regard to the magnitude of predation of adult female *T. turkestanii*, with the ladybird being more effective on cowpea than on white bean and red bean. The effect of host plants on the functional response of predatory insects has been documented in the literature. Coll and Ridgway (1995) found that *Orius insidiosus* Say had a lower predation rate on *Frankliniella occidentalis* (Pergande) on tomato

plants than on bean and maize plants. They suggested that the presence of glandular trichomes on the surface of tomato leaves might hamper predator searching efficiency and also provide more refuges to protect the thrips against predator attacks. In another experiment, De Clercq et al. (2000) reported that *Podius nigrispinus* (Say) had a lower feeding rate on beet armyworm, *Spodoptera exigua* (Hubner), on tomato than on sweet pepper and eggplant. They proposed that the glandular

trichomes on the plant surface and allelochemicals interference with the performance of the predator on the tomato resulted in impaired searching efficiency and predation ability. Karami Jamour and Shishehbor (2012) also reported that lower predation efficiency of *S. gilvifrons* on adult females of *T. turkestanii* on cucumber than on cowpea and castor bean leaves might be the result of, among other reasons, the dense trichomes covering the lower surface of cucumber leaves, which might decrease the movement of the predator and increase the prey handling time. Furthermore, the negative effect of leaf hairs and trichomes on the searching ability of predators and parasitoids has been reported by other researchers (Price et al., 1980; van Haren et al., 1987; Walter and O'Dowd 1992; Sütterlin and van Lenteren, 1997; Ricci and Capelletti, 1998; Croft et al., 1999), either as a mechanical mechanism slowing down the movements of the natural enemy (Price et al., 1980; Sütterlin and van Lenteren, 1997) or as adhesive secretions, which make the natural enemy spend a large amount of its time cleaning itself (van Haren et al., 1987).

The results obtained in the current study indicated that not only the host plant and prey density significantly affected the functional response of *S. gilvifrons* against adult females of *T. turkestanii*, but also there is a significant interaction between the host plant species and prey density on the functional response of *S. gilvifrons* preying on *T. turkestanii*. In this manner, the influence of the host plant becomes more obvious at high prey densities. A similar trend has been reported for *S. gilvifrons* fed on protonymphs and deutonymphs of *T. turkestanii* on three different host plant species (Karami Jamour and Shishehbor, 2012) and for the phytoseiid mite *Phytoseiulus persimilis* Athias-Henriot fed on *Tetranychus urticae* Koch eggs on three different plant species (Skirvin and Fenlon, 2001).

Prevez and Omkar (2005) stated that the handling time and attack rate are two parameters used to ascertain the magnitude of functional response. The handling time is a good index of the

consumption rate and predator effectiveness because it determines the accumulative time spent on identifying, chasing, capturing, killing, and eating the prey (Holling, 1959, 1965, 1966). Handling time may be influenced by different factors like the speed of the predator, prey movement, and the time spent overcoming individual prey (Hassell, 1978), which may be related to behavioral and physical mechanisms of prey defense (Ali et al., 2011). In the current study, the handling time recorded for *T. turkestanii* adult females on white beans was higher than that on red beans. The differences in handling times of *S. gilvifrons* in this study could be related to the differences observed in the density of leaf trichomes on the tested host plants. Leaf density was higher on white beans than that on red beans. Dense trichomes on white bean leaves can reduce the mobility of *Stethorus punctillum* (Weise) adults foraging for *Tetranychus urticae* Koch eggs but cannot affect the movement of mobile stages of *T. urticae* in the laboratory (Riddick and Wu, 2011). Host plant trichomes can reduce the predation potential of predators (Price et al., 1980; Walter and O'Dowd, 1992; Ricci and Capelletti, 1998; Riddick and Simmons, 2014).

O'Neil (1989), Wiedenmann and O'Neil (1991), and Messina and Hanks (1998) criticized the functional response studies in small laboratories. They argued that under the field conditions, the factors such as large searching areas, host plants, multispecies prey and/or predator systems, and weather conditions could influence the effectiveness of natural enemies. In the present study, for instance, the experiments were conducted in arenas, where no or very little spider mite webbing was produced. In the bean fields, all *T. turkestanii* stages are surrounded by webbing, which may limit the predation efficiency of *S. gilvifrons*. Thus, additional studies evaluating the effect of *S. gilvifrons* on *T. turkestanii* in field trials are recommended.

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REFERENCES

- Afshari, G.A. 1999. A survey on the ladybirds belong to genus *Stethorus* and study on the biology, prey consumption and population dynamics of *Stethorus gilvifrons* in sugarcane farms in Khuzestan, Iran. M. Sc, Thesis, Shahid Chamran University of Ahvaz, Ahvaz, Iran.
- Ahn, J.J., Kim, K.W., and Lee, J. H. 2010. Functional response of *Neoseiulus californicus* (Acari: Phytoseiidae) to *Tetranychus urticae* (Acari: Tetranychidae) on strawberry leaves. *Journal of Applied Entomology*, 134: 96- 104.
- Aksit, T., Cakmak, I., and Ozer, G. 2007. Effect of temperature and photoperiod on development and fecundity of acarophagous ladybird beetle, *Stethorus gilvifrons*. *Phytoparasitica*, 35(4): 357-366.
- Ali, M.P., Naif, A.A., and Huang, D. 2011. Prey consumption and functional response of a phytoseiid predator, *Neoseiulus womersleyi*, feeding on spider mite, *Tetranychus macfarlanei*. *Journal of Insect Science*, 11: 1- 11.
- Allahyari H., Azmayeshfard P., Nozari J. 2004. Effects of host on functional response of offspring in two populations of *Trissolcus grandis* on the sunn pest .*Journal of Applied Entomology*, 128: 39- 43.
- Biddinger D.J, Weber D.C., and Hull L.A. 2009. Coccinellidae as predator of mites: Stethorini in biological control. *Biological Control*, 51:268-283.
- Bolland H.R., Gutierrez J., and Flechtmann C.H.W. 1998. World catalogue of the spider mite family (Acari: Tetranychidae), with reference to taxonomy, synonymy, host plants and distribution. Brill academic Publisher Leiden. pp. 392.
- Carey, J.R., and J.W. Bradley, 1982. Developmental rates, vital schedules, sex ratios, and life tables for *Tetranychus urticae*, *Tetranychus turkestani* and *Tetranychus pacificus* (Acarina: Tetranychidae) on cotton. *Acarologia*, 23 (4): 333-345.
- Cedola, C.V., Sanchez, N.E., and Liljestrom, G.G. 2001. Effect of tomato leaf hairiness on functional and numerical response of *Neoseiulus californicus* (Acari: Phytoseiidae). *Experimental and Applied Acarology*, 25: 819- 831.
- Chazeau, J. 1985. Predaceous insects. In: Helle, W. and Sabelis, M.W. (editors) *World crop pests, spider mites: Their biology, natural enemies and control*. Elsevier Publisher, Amsterdam, IB: pp: 211-246.
- Coll, M., and Ridgway, R.L. 1995. Functional and numerical responses of *Orius insidiosus* (Heteroptera: Anthracoridae) to its prey in different vegetable crops. *Annals of the Entomological Society of America*, 88: 732-738.
- Croft, P., Fenlon, J., Jacobson, R.J., and Dubas, J. 1999. Effect of tomato conditioning on *Phytoseiulus persimilis* Athias- Henriot (Acari: Phytoseiidae) population growth. *IOBC WPRS Bulletin*, 22(1), 45–48.
- De Clercq, P., Mohaghegh, J., and Tirry, L. 2000. Effect of Host Plant on the functional response of the predator *Podisus nigrispinus* (Heteroptera: Pentatomidae). *Biological Control*, 18: 66-70.

- Dicke, M., Sabelis, M.W., Takabayashi, J., Bruin, J., Posthumus, M.A. 1990. Plant strategies of manipulating predator prey interactions through allelochemicals: prospects for application in pest control. *Journal of Chemical Ecology*, 16, 3091- 3018.
- Eisner, T. 1970. Chemical defense against predation in arthropods. In Sondheimer, E., Simeone, J. B. (eds). *Chemical ecology*. Academic Press, New York, New York, USA. Pp: 157-217.
- Farhadi, R., Allahyari, H., Juliano, S.A., 2010. Functional response of larval and adult stages of *Hippodamia variegata* (Coleoptera: Coccinellidae) to different species of *Aphis fabae* (Hemiptera: Aphididae). *Environmental Entomology*, 39 (5): 1586- 1592.
- Gontijo, L.M., Nechols, J.R., Margolies, D.C., and Cloyd, R.A. 2012. Plant architecture and prey distribution influence foraging behavior of the predatory mite *Phytoseiulus persimilis* (Acari: Phytoseiidae). *Experimental and Applied Acarology*, 56: 23- 32.
- Haji-zadeh, G. Kamali K., and Baiat|asadi, H. 1993. Investigations on the functional response and population fluctuation of *Stethorus gilvifrons* on red spider mite, *Panonychus ulmi* in orchards around Karaj, Iran. *Pest and Disease Journal*, 61: 117- 131.
- Haque, M.F., Islam, M.W., Khalequzzaman, M. 2010. Functional response of *Phytoseius persimilis* Athias-Henriot to *Tetranychs urticae* Koch: effect of prey life stage and temperature. *University Journal of Zoology, Rajshahi University*, 29 (1): 1- 8.
- Hassell, M.P. Lawton, J.H., and Beddington, J.R. 1977. Sigmoid functional response by invertebrate predators and parasitoids. *Journal of Animal Ecology*, 46: 249- 262.
- Hassell, M. 1978. *The dynamics of orthropod predator – prey system*. Princeton University Press, Princeton. 237 pp.
- Helle, W., and Sabelis, M.W. 1985. *Spider mites: Their biology, natural enemies, and control*. Vol. 1A and1B. Amsterdam, Elsevier.
- Holling, C.S. 1959. Some characteristics of simple types of predation and parasitism. *Canadian Entomologist*, 91: 385- 398.
- Holling, C.S. 1965. The functional response of predators to prey density and its role in mimicry and population regulation. *Memory of the Entomological Society of Canada*, 48: 3- 60.
- Holling, C.S. 1966. The functional response of invertebrate predators to prey density. *Memory of the Entomological Society of Canada*, 98: 5- 86.
- Hoy, M.A. 2011. *Agricultural acarology: Introduction to integrated mite management*. CRC Press. Taylor & Francis Group.
- Imani, Z., Shishehbor, P., and Mossadegh, M.S. 2010. Functional response of the coccinellid predator *Stethorus gilvifrons* on different stages of *Eutetranychus orientalis* (Klein). *Plant Protection (Journal of Agricultural Science)*, 33 (1): 1- 10.
- Jeppson, L.R., Keifer, H.H., and Baker, E.W. 1975. *Mites injurious to economic plants*. University of California Press. 614 pp.

- Juliano, S.A. 2001. Nonlinear curve fitting: predation and functional response curves. In: Scheiner, S. M., Gurevitch, J. (eds) Design and analysis of ecological experiments. New York, Oxford University Press, pp: 178- 196.
- Kamali, K., Ostowan, H., and Atamehr, A. 2001. A catalogue of mites and ticks (Acari) of Iran. Tehran, Islamic Azad University Scientific Publication Centre, pp. 192.
- Karami Jamour, T., Shishebor, P., and Mossadegh, M.S. 2012. Biology and life table parameters of strawberry spider mite, *Tetranychus turkestanii* studied on three cucurbit plants. Iranian Plant Protection Sciences Journal, 43 (1): 23- 31.
- Karami Jamour, T., and Shishebor, P. 2012. Host plant effects on the functional response of *Stethorus gilvifrons* to strawberry spider mites. Biocontrol Science and Technology, 22: 101-110.
- Khanamani, M., Fathipour, Y., Talebi, A.A., Mehrabadi, M. 2017. Quantitative analysis of long-term mass rearing of *Neoseiulus californicus* (Acari: Phytoseiidae) on almond pollen. Journal of Economic Entomology, 110 (4): 1442- 1450.
- Kohpayezadeh Isfahani, P., and Mossadegh, M.S. 1993. Part of the coccinellids (Col.: Coccinellidae) fauna of Kerman province. Plant Protection (Scientific Journal of Agriculture), 16 (1, 2): 64- 75.
- Kolberg, I., Bylund, H., Jonsson, T., Schmidt, A., Greshenzon, J., Bjorkman, C. 2015. Temperature affects insect outbreak risk through tritrophic interactions mediated by plant secondary compounds. Ecosphere, 6 (6): 102.
- Li, D.X., Tian, J., Shen, Z.R. 2007. Functional response of the predator Scolothrips takahashii to hawthorn spider mite, *Tetranychus viennensis*: effect of age and temperature. BioControl, 52: 41- 61.
- Matin, M. 2007. Biology and predation of ladybird *Stethorus gilvifrons* on old world date mite, *Oligonychus afrasiaticus* McGregor. M. Sc. Thesis, Mohaghegh Ardabili University, Ardabil, Iran.
- McMurtry, J., Huffaker, C.B., and Dev Rie, M.V. 1970. Ecology of *Tetranychus*. In: enemies: Their biological characters and impact of spray practices. Hilgardia, 40: 331-390.
- Mehr-khou, F. 2006. Searching behavior of *Stethorus gilvifrons* on two spotted spider mite, *Tetranychus urticae*. M. Sc. Thesis, Tarbiat Modares University, Tehran, Iran.
- Messina, F.J., and Hanks, J.B. 1998. Host plant alters the shape of functional response of an aphid predator (Coleoptera: Coccinellidae). Environmental Entomology, 27: 1196- 1202.
- Modarres Awal, M. 2012. List of agricultural pests and their natural enemies in Iran. Revised Edition. Ferdowsi University of Mashhad Publication.
- Mossadegh, M.S., and Kocheili, F. 1993. Some of the ladybirds (Coccinellidae) fauna of Khuzestan, southern Iran. Plant Protection (Scientific Journal of Agriculture). 16 (1, 2): 23- 30.
- Mossadegh M.S., and Kocheili F. 2003. A semi descriptive checklist of identified species of arthropods (Agricultural, Medical, ...) and other pests from Khuzestan, Iran. Shahid Chamran University Press, Ahvaz, 342 pp

- Murdoch W.W. 1969. Switching in general predators: experiments on predator specificity and stability of prey populations. *Ecological Monographs*, 39 (4): 335–364.
- Nemati, A, Soleiman Nejadian, E., Shishehbor, P., and Kamali, K. 2005. Evaluation of the effect of temperature on biological parameters of two spotted spider mite *Tetranychus turkestanii* Ugarov and Nikolski (Acari: Tetranychidae). *Plant Protection (Scientific Journal of Agriculture)*, 28 (1): 209- 222.
- O’Neil, R.J. 1989. Comparison of laboratory and field measurements of the functional response of *Podisus maculiventris* (Heteroptera: Pentatomidae). *Journal of Kansas Entomological Society*, 62: 148-155.
- Prevez, A., and Omkar, P. 2005. Functional responses of coccinellid predators: an illustration of a logistic approach. *Journal of Insect Science*, 5: 1- 6.
- Price, P.W., Bouton, C. E., Gross, P., McPheron, B.A., Thompson, J.N., and Weis, A.E. 1980. Interaction among tritrophic levels: influence of plant on interaction between herbivores and natural enemies. *Annual Review of Ecology and Systematics*, 11: 41- 56.
- Putman, W.L. 1955. Bionomics of *Stethorus punctillum* Weise (col: Coccinellidae) in Ontario. *Canadian Entomologist*, 87:33-9.
- Ricci, C. and Capelletti, G. 1998. Relationship between some morphological structures and locomotion of *Clithostethus arcuatus* Rossi (Coleoptera; Coccinellidae), a whitefly predator. *Frustula Entomologica*, 11, 195–202.
- Riddick, E.W., Rojas, M.G., and Wu, Z. 2011. Lima bean-lady beetle interactions: spider mite mediates sub lethal effects of its host plant on growth and development of its predator. *Arthropod-Plant Interactions*, 5: 287-296.
- Riddick, E.W., and Wu, Z. 2011. Lima bean–lady beetle interactions: hooked trichomes affect survival of *Stethorus punctillum* larvae. *BioControl*, 56: 55–63.
- Riddick, E. W., and Simmons, A. 2014. Do plant trichomes cause more than good to predatory insects? *Pest Management Sciences*, 70: 1655- 1665.
- Rogers, D. 1972. Random search and insect population models. *Journal of Animal Ecology*, 369- 383.
- SAS Institute., 2012. Version 9.1, Qualification Tools user Guide, NC, USA: Cary.
- Skirvin, D.J., and Fenlon, J.S. 2001. Plant species modifies the functional response of *Phytoseiulus persimilis* (Acari: Phytoseiidae) to *Tetranychus urticae* (Acari: Tetranychidae): implications for biological control. *Bulletin of Entomological Research*, 91:61-67.
- Skirvin, D.J., and Fenlon, J.S. 2003. The effect of temperature on the functional response of *Phytoseiulus persimilis* (Acari: Phytoseiidae). *Experimental and Applied Acarology*, 31: 37- 49.
- Sohrabi, F., and Shishehbor, P. 2007. Functional and numerical response of *Stethorus Gilvifrons* Mulsant (Col.: Coccinellidae) feeding on *Tetranychus turkestanii* Ugarov and Nikolski (Acari: Tetranychidae). *Pakistan Journal of Biological Sciences*, 10 (24): 4563-4566.

- Sohrabi, F., and Shishehbor, P. 2008. Effects of host plant and temperature on growth and reproduction of the strawberry spider mite *Tetranychus turkestanii* Ugarov & Nikolski (Acari: Tetranychidae). *Systematic and Applied Acarology*, 13: 26-32.
- Solomon, M. E. 1949. The natural control of animal populations. *Journal of Animal Ecology*, 18: 1- 35.
- Sütterlin, S., and van Lenteren, J.C. 1997. Influence of hairiness of *Gerbera jamesonii* leaves on the searching efficiency of the parasitoid *Encarsia formosa*. *Biological Control*, 9: 157- 165.
- Takafuji, A., and Deguchi, K. 1980. Functional responses of a predacious phytoseiid mite in different sizes of experimental universe. *Applied and Experimental Zoology*, 15: 355- 357.
- Tomeczyk A., and Kropczynska, D. 1985. Effects of host plants. In: Helle W. Sabelis M. W. (eds). *Spider mites: their biology, natural enemies and control*. Volume 1A. Elsevier Science Publisher. Pp. 317- 330.
- Van Haren, R.J.F., Steenhuis, M.M., Sabelis, M.W., and De Ponti, O.M.B. 1987. Tomato stem trichomes and dispersal success of *Phytoseiulus persimilis* relative to its prey *Tetranychus urticae*. *Experimental and Applied Acarology*, 3: 115- 121.
- Walter, D.E., and O'Dowd, D.J. 1992. Leaf morphology and predators: effect of leaf domatia on the abundance of predatory mites (Acari: Phytoseiidae). *Environmental Entomology*, 21: 478- 484.
- Wiedenmann, R. N. and O'Neil, R. J. 1991. Searching behavior and time budgets of the predator *Podisus maculiventris*. *Entomologia Experimentalis et Applicata*, 60: 83- 93.
- Zarghami, S., Mossadegh, M.S., Kocheili, F., Allahyari, H., Rasekh, A. 2016. Functional responses of *Nephus arcuatus* Kapur (Coleoptera: Coccinellidae), the most important predator of spherical mealybug *Nipaecoccus viridis* (Newstead). *Psyche*, 2016: 1-9.
- Zhang, Z. 2003. *Mites of greenhouses identification, biology and control*. CABI Publishing, 234 pp.



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اثر گیاهان میزبان بر واکنش تابعی کفشدوزک کنه خوار *Stethorus gilvifrons* روی کنه *Tetranychus turkestanii*

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چکیده

کنه تارتین توت فرنگی *Tetranychus turkestanii* Ugarov and Nikolski یکی از مهم ترین آفات گیاهان زراعی و گلخانه ای در خوزستان است. هدف این مطالعه بررسی واکنش تابعی کفشدوزک *Stethorus gilvifrons* Mulsant روی تراکم های مختلف کنه *T. turkestanii* روی سه نوع لوبیا: لوبیا چشم بلبلی (*Vigna unguiculata* L.)، لوبیا سفید (*Phaseolus lunatus* L.) و لوبیا قرمز (*Phaseolus calcaratus* Roxb.) در دمای 2 ± 30 درجه سلسیوس، رطوبت نسبی 10 ± 60 درصد و دوره روشنائی: تاریکی ۸:۱۶ ساعت بود. هر کفشدوزک بالغ به مدت ۲۴ ساعت بر روی برگ های هر کدام از میزبان های گیاهی مذکور حامل تراکم هایی از ۸، ۱۲، ۲۴، ۴۸ و ۹۶ کنه بالغ *T. turkestanii* روی دیسک برگی در یک پتری دیش با قطر ۹ سانتی متر نگهداری شد. تجزیه و تحلیل رگرسیونی لوجستیک نشان داد نوع واکنش تابعی روی لوبیا چشم بلبلی، لوبیا سفید و لوبیا قرمز به ترتیب نوع سوم، دوم و دوم بود. بیش ترین نرخ حمله (۰/۱۲۱۹) و کوتاه ترین زمان دست یابی (۰/۲۳۰۱) به ترتیب روی لوبیا سفید و لوبیا قرمز ثبت شد. بیش ترین نرخ شکارگری (T/T_h) روی گیاهان مذکور ۰/۷۲/۷۴، ۰/۶۰/۴۲ و ۰/۱۰۴/۳۰ کنه در روز بود. این مطالعه آزمایشگاهی پیشنهاد می کند که خصوصیات گیاه میزبان بر توانایی کفشدوزک *S. gilvifrons* جهت واکنش به تغییر در تراکم کنه *T. turkestanii* اثر می گذارد.

کلیدواژه ها: نرخ حمله، زمان دست یابی، بقولات، کنه تارتین توت فرنگی